

Non contiguous-finished genome sequence and description of *Cellulomonas massiliensis* sp. nov.

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Cellulomonas massiliensis strain JC225^T sp. nov. is the type strain of *Cellulomonas massiliensis* sp., a new species within the genus *Cellulomonas*. This strain, whose genome is described here, was isolated from the fecal flora of a healthy Senegalese patient. *C. massiliensis* is an aerobic rod-shaped bacterium. Here we describe the features of this organism, together with the complete genome sequence and annotation. The 3,407,283 bp long genome contains 3,083 protein-coding and 48 RNA genes.

Introduction

Cellulomonas massiliensis strain JC225^T (= CSUR P160 = DSM 25695) is the type strain of *C. massiliensis* sp. nov. This bacterium is a motile, Gram-positive, aerobic, indole-negative rod that was isolated from the stool of a healthy Senegalese patient as part of a culturomics study aiming at cultivating all bacterial species within human feces [1].

The current approach to the classification of prokaryotes, known as polyphasic taxonomy, relies on a combination of phenotypic and genotypic characteristics [2]. However, as more than 3,000 bacterial genomes have been sequenced [3], and proteomic information is more becoming more readily accessible [4], we recently proposed that genomic information should be integrated in the description of new bacterial species [5-11].

The genus *Cellulomonas* was created in 1923 to reclassify several bacteria previously classified as *Bacillus* species [12]. To date, this genus is made of 19 species [13-24]. The two species that are the most phylogenetically related to *C. massiliensis* are *C. composti* [17] and *C. persica* [21]. Most of these species were originally isolated from environmental samples, notably from habitats enriched in cellulose, such as soil or sugar fields, and occasionally from the rumen and activated sludge. Rare cases of human endocarditis [25], osteomyelitis [25], endophthalmitis [26] and cholecystitis [27] caused by *Cellulomonas* species have been reported. To

date, members of the genus *Cellulomonas* have not been described in the normal fecal flora.

Here we present a summary classification and a set of features for *C. massiliensis* sp. nov. strain JC225^T together with the description of the complete genomic sequencing and annotation. These characteristics support the circumscription of the species *C. massiliensis*.

Classification and features

A stool sample was collected from a healthy 16-year-old male Senegalese volunteer patient living in Dielmo (a rural village in the Guinean-Sudanian zone in Senegal), who was included in a research protocol. Written assent was obtained from this individual; no written consent was needed from his guardians for this study because he was older than 15 years old (in accordance with the previous project approved by the Ministry of Health of Senegal and the assembled village population and as published elsewhere [28]).

Both this study and the assent procedure were approved by the National Ethics Committee of Senegal (CNERS) and the Ethics Committee of the Institut Fédératif de Recherche IFR48, Faculty of Medicine, Marseille, France (agreement numbers 09-022 and 11-017). Several other new bacterial species were isolated from this specimen using various culture conditions, including the recently described *Anaerococcus senegalensis*, *Bacillus*

timonensis, *Alistipes senegalensis*, *Alistipes timonensis*, *Clostridium senegalense*, *Paenibacillus senegalensis* and *Peptoniphilus timonensis* [5-11], thus suggesting that the human digestive flora is far from being fully known. The fecal specimen was preserved at -80°C after collection and sent to Marseille. Strain JC225 (Table 1) was isolated in May 2011 by passive filtration of the stool and aerobic incubation on Brain Heart Infusion agar at 37°C. This strain exhibited a nucleotide sequence similarity of 98.3% with *Cellulomonas composti* (Kang et al 2007), the phylogenetically closest validated *Cellulomonas* species (Figure 1) that was

cultivated from cattle farm compost [17]. This value was lower than the 98.7% 16S rRNA gene sequence threshold recommended by Stackebrandt and Ebers to delineate a new species without carrying out DNA-DNA hybridization [39]. By comparison to the Genbank database [40] strain JC225^T also exhibited a nucleotide sequence similarity greater than 99.5% with *Cellulomonas* sp. strain 3335BRRJ isolated from clean room environments (Genbank accession number FJ200382). This bacterium is most likely classified within the same species as strain JC225^T (Figure 1).

Table 1. Classification and general features of *Cellulomonas massiliensis* strain JC225^T

MIGS ID	Property	Term	Evidence code ^a
	Current classification	Domain <i>Bacteria</i>	TAS [29]
		Phylum <i>Actinobacteria</i>	TAS [30]
		Class <i>Actinobacteria</i>	TAS [31]
		Order <i>Actinomycetales</i>	TAS [31-34]
		Family <i>Cellulomonadaceae</i>	TAS [31,34-37]
		Genus <i>Cellulomonas</i>	TAS [12,32]
		Species <i>Cellulomonas massiliensis</i>	IDA
		Type strain JC225 ^T	IDA
	Gram stain	positive	IDA
	Cell shape	rod	IDA
	Motility	positive	IDA
	Sporulation	nonsporulating	IDA
	Temperature range	mesophilic	IDA
	Optimum temperature	37°C	IDA
MIGS-6.3	Salinity	growth in BHI medium + 5% NaCl	IDA
MIGS-22	Oxygen requirement	aerobic	IDA
	Carbon source	galactose	NAS
	Energy source	chemoorganotrophic	NAS
MIGS-6	Habitat	human gut	IDA
MIGS-15	Biotic relationship	free living	IDA
	Pathogenicity	unknown	NAS
	Biosafety level	2	
MIGS-14	Isolation	human feces	
MIGS-4	Geographic location	Senegal	IDA
MIGS-5	Sample collection time	September 2010	IDA
MIGS-4.1	Latitude	13.7167	IDA
MIGS-4.1	Longitude	- 16.4167	IDA
MIGS-4.3	Depth	surface	IDA
MIGS-4.4	Altitude	51 m above sea level	IDA

Evidence codes - IDA: Inferred from Direct Assay; TAS: Traceable Author Statement (i.e., a direct report exists in the literature); NAS: Non-traceable Author Statement (i.e., not directly observed for the living, isolated sample, but based on a generally accepted property for the species, or anecdotal evidence). These evidence codes are from the Gene Ontology project [38]. If the evidence is IDA, then the property was directly observed for a live isolate by one of the authors or an expert mentioned in the acknowledgements.

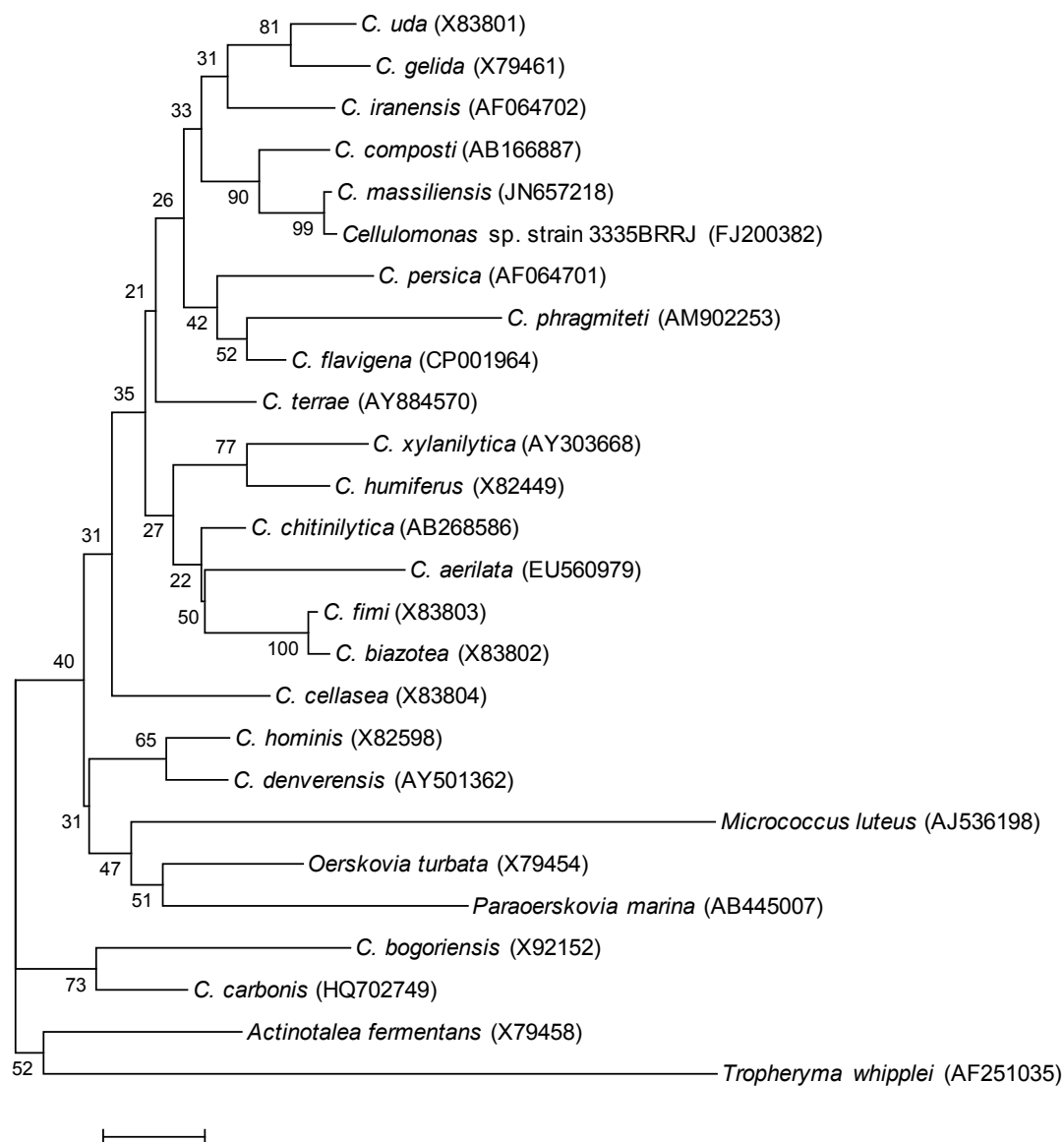


Figure 1. Phylogenetic tree highlighting the position of *Cellulomonas massiliensis* strain JC225 relative to other type strains within the genus *Cellulomonas* and other members of the family *Cellulomonadaceae*. GenBank accession numbers are indicated in parentheses. Sequences were aligned using CLUSTALW, and phylogenetic inferences obtained using the maximum-likelihood method within the MEGA software. Numbers at the nodes are bootstrap values obtained by repeating the analysis 500 times to generate a majority consensus tree. The scale bar indicates a 1% nucleotide sequence divergence.

Different growth temperatures (25, 30, 37, 45°C) were tested; no growth occurred at 25°C or 45°C, growth occurred between 30 and 37°C, and optimal growth was observed at 37°C. Colonies were transparent and smooth with a diameter of 1 mm on blood-enriched Columbia agar and Brain Heart Infusion (BHI) agar. Growth of the strain was tested under anaerobic and microaerophilic conditions using GENbag anaer and GENbag microaer systems, respectively (BioMérieux), and in the presence of air, with or without 5% CO₂. Optimal

growth was achieved aerobically. Weak growth was observed under microaerophilic condition and with 5% CO₂. No growth was observed under anaerobic conditions. Gram staining showed Gram-positive rods. A motility test was positive. Cells grown on agar are Gram-positive (Figure 2), with a diameter and length ranging from 0.37 to 0.60 µm (mean, 0.48 µm), and from 0.55 to 1.4 µm (mean, 0.95 µm), respectively, in electron microscopy, (Figure 3).

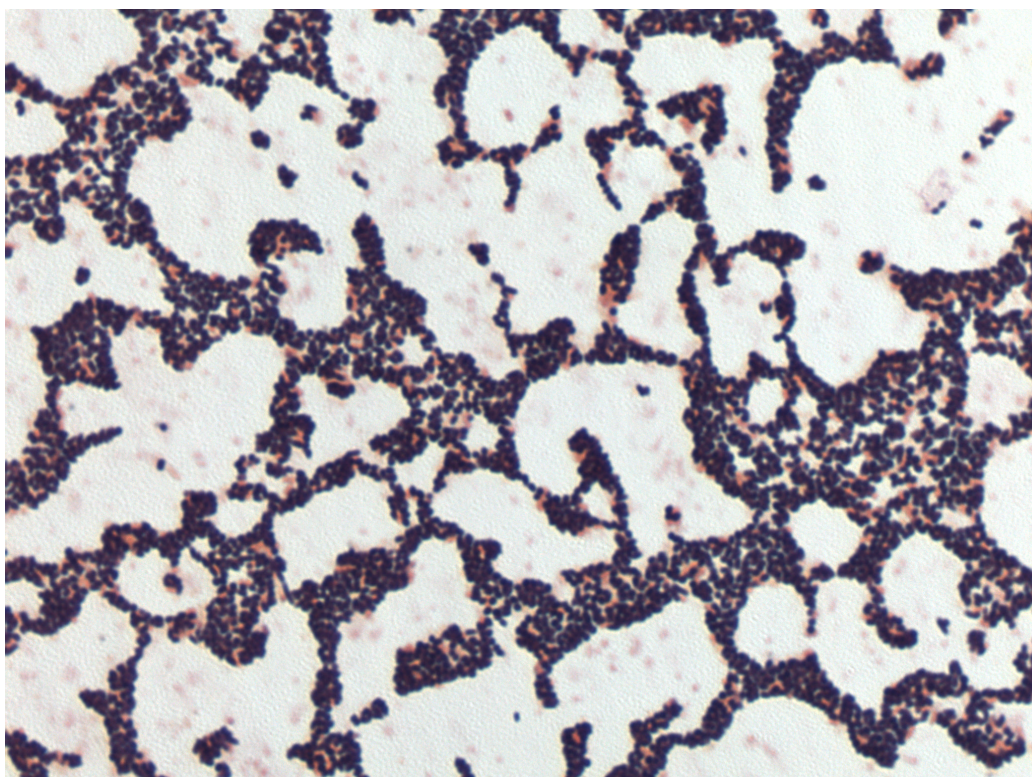


Figure 2. Gram staining of *C. massiliensis* strain JC225^T

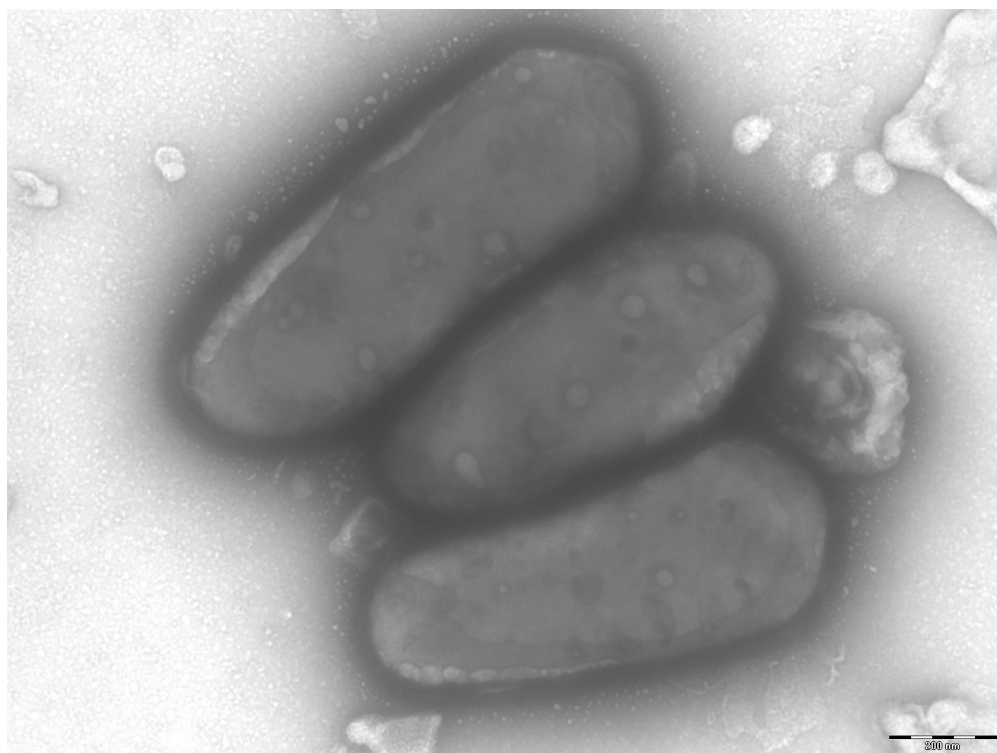


Figure 3. Transmission electron microscopy of *C. massiliensis* strain JC225^T, using a Morgani 268D (Philips) at an operating voltage of 60kV. The scale bar represents 200 nm.

Strain JC225^T exhibited catalase and oxidase activities. Using the API 20 NE system (BioMérieux), a positive reaction was obtained for aesculin hydrolysis and β -galactosidase. Negative reactions were obtained for nitrate reduction, indole production, glucose fermentation, arginine dihydrolase, urease, gelatin hydrolysis, and glucose, arabinose, mannose, mannitol N-acetylglucosamine, maltose, gluconate, caprate, adipate,

malate, citrate, and phenyl-acetate assimilation. *C. massiliensis* is susceptible to amoxicillin, imipenem, gentamicin, and ciprofloxacin but resistant to trimethoprim/sulfamethoxazole and metronidazole. By comparison to *C. composti* [17], *C. massiliensis* differed in motility, nitrate reduction, gelatine hydrolysis, carbohydrate assimilation, and catalase activity (Table 2).

Table 2. Differential phenotypic characteristics of five *Cellulomonas* strains[†].

Properties	<i>C. massiliensis</i>	<i>C. composti</i>	<i>C. persica</i>	<i>C. flavigena</i>	<i>C. iranensis</i>
Oxygen requirement	aerobic	Facultative anaerobic	aerobic	aerobic	aerobic
Gram stain	+	+	+	+	+
Motility	+	–	+	–	+
Production of					
Catalase	+	–	na	+	na
Nitrate reductase	–	+	+	+	+
Urease	–	–	+	–	+
β -galactosidase	+	na	na	na	na
N-acetyl-glucosamine	–	na	na	na	na
Arginine dihydrolase	–	na	na	na	na
Fermentation for					
Sucrose	–	+	na	+	na
Glucose	–	+	na	+	na
Mannitol	–	–	na	–	na
Gluconate	–	–	na	W	na
Maltose	–	+	na	+	na
Hydrolysis of					
Gelatin	+	w	w	+	w
Esculin	–	+	na	+	na
G+C content (mol%)	71.2	73.7	na	72.7-74.8	na
Habitat	Human gut	Compost	Forest humus soil	Soil	Forest humus soil

[†]*Cellulomonas massiliensis* strain JC225^T, *Cellulomonas composti* strain TR7-06^T, *Cellulomonas persica* strain I^T, *Cellulomonas flavigena* strain DSM 20109^T, and *Cellulomonas iranensis* strain O^T.

na = data not available; w = weak

na = data not available; w = weak

Matrix-assisted laser-desorption/ionization time-of-flight (MALDI-TOF) MS protein analysis was carried out as previously described [5,41] using a Microflex spectrometer (Bruker Daltonics, Germany). Twelve distinct deposits were done for strain JC225 from 12 isolated colonies. The 12 JC225 spectra were imported into the MALDI BioTyper software (version 2.0, Bruker) and analyzed by standard pattern matching (with default parameter settings) against the main spectra of

3,769 bacteria, which were used as reference data in the BioTyper database. The database contained 11 spectra from 8 validly published *Cellulomonas* species, including *Cellulomonas composti*, the phylogenetically closest species to *C. massiliensis*. No significant score was obtained for strain JC225^T, thus suggesting that our isolate was not a member of a known species within the Bruker database. We incremented our database with the reference spectrum from strain JC225^T (Figure 4).

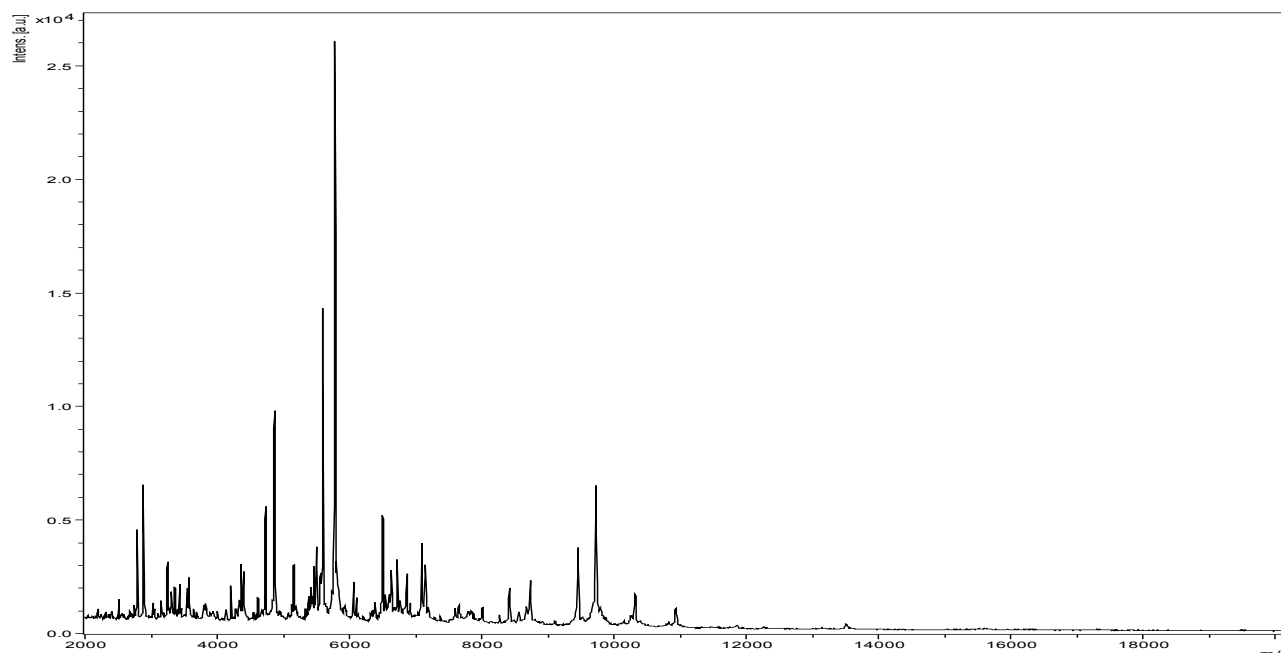


Figure 4: Reference mass spectrum from *C. massiliensis* strain JC225^T. Spectra from 12 individual colonies were compared and a reference spectrum was generated.

Genome sequencing information

Genome project history

The organism was selected for sequencing on the basis of its phenotypic differences, phylogenetic position and 16S rRNA similarity to other members of the genus *Cellulomonas* and is part of a study of the human digestive flora aiming at isolating all bacterial species within human feces. It was the fourth genome of a *Cellulomonas* species and the first genome of *Cellulomonas massiliensis* sp. nov. The EMBL accession number is CAHD00000000 and consists of 250 contigs (≥ 200 bp). Table 3 shows the project information and its association with MGS version 2.0 compliance [42].

Growth conditions and DNA isolation

Cellulomonas massiliensis sp. nov. JC225^T (= CSUR P160 = DSM 25695) was grown aerobically on 5% sheep blood-enriched Columbia agar (BioMérieux) at 37°C. Ten petri dishes were spread and resuspended in 3×100µl of G2 buffer (EZ 1 DNA Tissue kit, Qiagen). A first mechanical lysis was performed using glass powder on a Fastprep-24 device (MP Biomedicals, Illkirch, France) during 2×20 seconds. DNA was then treated with 2.5µg/µL lysozyme (30 minutes at 37°C) and extracted using a BioRobot EZ 1 Advanced XL (Qiagen). The DNA was then concentrated and purified using a Qiaamp kit (Qiagen). The yield and the concentration were measured using a Quant-it Picogreen kit (Invitrogen) on a Genios_Tecan fluorometer at 78.9 ng/µl.

Table 3. Project information

MIGS ID	Property	Term
MIGS-31	Finishing quality	High-quality draft
MIGS-28	Libraries used	Paired-end 3 Kb library
MIGS-29	Sequencing platforms	454 GS FLX Titanium
MIGS-31.2	Fold coverage	25×
MIGS-30	Assemblers	Newbler version 2.5.3
MIGS-32	Gene calling method	Prodigal
	EMBL ID	CAHD000000000
	EMBL Date of Release	May 30, 2012
	Project relevance	Study of the human gut microbiome

Genome sequencing and assembly

Both shotgun sequencing and paired-end sequencing strategies were used (Roche). Both libraries were pyrosequenced on a GS FLX Titanium sequencer (Roche). This project was loaded onto a single 1/4 region of a PTP Picotiterplate (Roche, Meylan, France) for the shotgun library and 2 × 1/4 region for the 3-kb paired-end library. The shotgun library was constructed with 500 ng of DNA with the GS Rapid library Prep kit as described by the manufacturer (Roche). For the paired-end library, 5 µg of DNA was mechanically fragmented on a Hydroshear device (Digilab, Holliston, MA, USA) with an enrichment size at 3–4 kb. The DNA fragmentation was visualized through an Agilent 2100 BioAnalyzer on a DNA labchip 7500 with an optimal size of 3.216 kb. The library was constructed according to the 454 Titanium paired-end protocol (Roche). Circularization and nebulization were performed and generated a pattern with an optimum at 395 bp. After PCR amplification through 17 cycles followed by double size selection, the single stranded paired-end library was quantified on with a Quant-it Ribogreen kit (Invitrogen) on a Genios Tecan fluorometer at 132 pg/µL. The library concentration equivalence was calculated at 6.11E+08 molecules/µL. The libraries were stored at -20°C until further use.

The shotgun library was clonally amplified with 3 cpb in 3 emPCR reactions and the 3-kb paired-end library was amplified with 0.5 cpb in 4 emPCR reactions with the GS Titanium SV emPCR Kit (Lib-L) v2 (Roche). The yield of the shotgun emPCR reactions was 10.13%, and the yields of the paired-end emPCRs was 8.6%, in the range of 5 to 20% from the Roche procedure.

Approximately 790,000 beads for both the shotgun and paired-end libraries were loaded on the GS Titanium PicoTiterPlate PTP Kit 70×75 and sequenced with the GS FLX Titanium Sequencing Kit XLR70 (Roche). The runs were performed over-

night and then analyzed on the cluster through the gsRunBrowser and Newbler Assembler (Roche). A total of 255,758 and 256,082 passed filter wells were obtained for the shotgun and paired-end strategies, respectively, and generated 86.75 and 78.45 Mb of DNA sequence with length averages of 339 and 313 bp, respectively. The filter passed sequences were assembled using Newbler with 90% identity and 40 bp as overlap. The final assembly identified 250 contigs (>200 bp) arranged into 5 scaffolds and generated a genome size of 3.40 Mb.

Genome annotation

Open Reading Frames (ORFs) were predicted using Prodigal [43] with default parameters but the predicted ORFs were excluded if they were spanned a sequencing GAP region. The predicted bacterial protein sequences were searched against the GenBank database [40] and the Clusters of Orthologous Groups (COG) databases using BLASTP. The tRNAscan-SE tool [44] was used to find tRNA genes, whereas ribosomal RNAs were found by using RNAmmer [45]. Transmembrane domains and signal peptides were predicted using TMHMM [46] and SignalP [47], respectively. ORFans were identified if their BLASTP *E*-value was lower than 1e-03 for alignment length greater than 80 amino acids. If alignment lengths were smaller than 80 amino acids, we used an *E*-value of 1e-05. Such parameter thresholds have been used in previous works to define ORFans. To estimate the mean level of nucleotide sequence similarity at the genome level between *C. massiliensis* and *C. flavigena* and *C. fimi* (EMBL accession numbers CP001964 and CP002666, respectively), the only two available genomes from validly published *Cellulomonas* species to date, we compared the ORFs only using BLASTN at a query coverage of ≥ 70% and a minimum nucleotide length of 100 bp.

Genome properties

The genome is 3,407,283 bp long (1 chromosome, but no plasmid) with a 71.22% G+C content (Table 4 and Figure 5). It is composed of 5 scaffolds. Of the 3,131 predicted genes, 3,083 were protein-coding genes, and 48 were RNAs (1 rRNA operon and 45 tRNA genes). A total of 2,184 genes

(70.84%) were assigned a putative function, and 256 genes were identified as ORFans (8.30%). The remaining genes were annotated as hypothetical proteins. The distribution of genes into COGs functional categories is presented in Table 5. The properties and the statistics of the genome are summarized in Table 4 and 5.

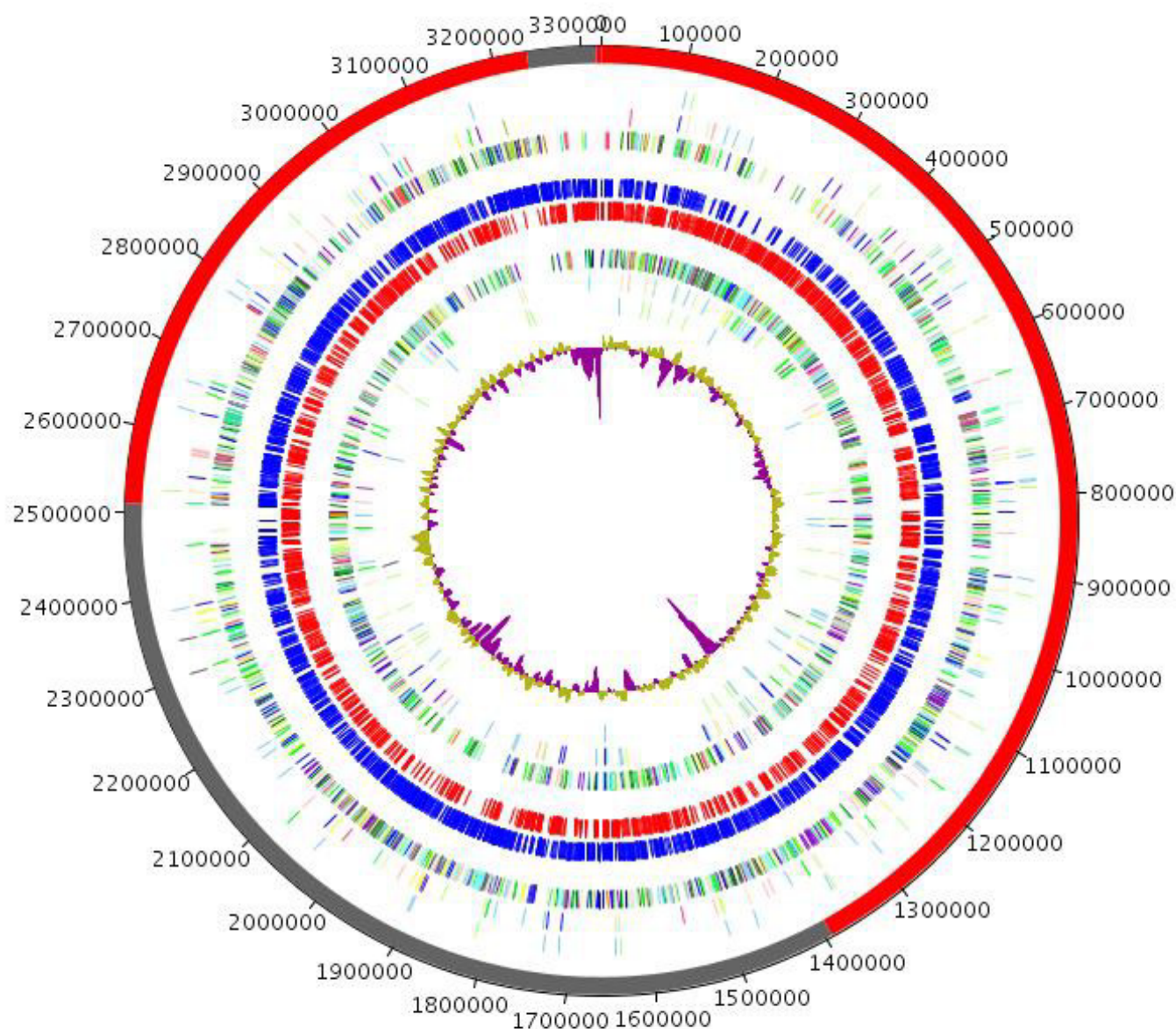


Figure 5. Graphical circular map of the *C. massiliensis* strain JC225^T genome. From outside to the center: scaffolds (red / grey), COG category of genes on the forward strand (three circles), genes on forward strand (blue circle), genes on the reverse strand (red circle), COG category on the reverse strand (three circles), G+C content.

Table 4. Nucleotide content and gene count levels of the genome

Attribute	Value	% of total ^a
Genome size (bp)	3,407,283	100
DNA coding region (bp)	3,091,035	90.72
DNA G+C content (bp)	2,426,733	71.22
Total genes	3,131	100
RNA genes	48	1.53
Protein-coding genes	3,083	98.47
Genes with function prediction	2,184	70.84
Genes assigned to COGs	2,155	69.9
Genes with peptide signals	387	12.55
Genes with transmembrane helices	721	23.39

The total is based on either the size of the genome in base pairs or the total number of protein coding genes in the annotated genome.

Table 5. Number of genes associated with the 25 general COG functional categories

Code	Value	%age	Description
J	140	4.54	Translation
A	1	0.03	RNA processing and modification
K	225	7.3	Transcription
L	110	3.57	Replication, recombination and repair
B	1	0.03	Chromatin structure and dynamics
D	20	0.65	Cell cycle control, mitosis and meiosis
Y	0	0	Nuclear structure
V	27	0.88	Defense mechanisms
T	116	3.76	Signal transduction mechanisms
M	111	3.6	Cell wall/membrane biogenesis
N	42	1.36	Cell motility
Z	0	0	Cytoskeleton
W	1	0.03	Extracellular structures
U	41	1.33	Intracellular trafficking and secretion
O	82	2.66	Posttranslational modification, protein turnover, chaperones
C	132	4.28	Energy production and conversion
G	253	8.21	Carbohydrate transport and metabolism
E	276	8.95	Amino acid transport and metabolism
F	68	2.21	Nucleotide transport and metabolism
H	85	2.76	Coenzyme transport and metabolism
I	68	2.21	Lipid transport and metabolism
P	112	3.63	Inorganic ion transport and metabolism
Q	42	1.36	Secondary metabolites biosynthesis, transport and catabolism
R	323	10.48	General function prediction only
S	175	5.68	Function unknown
-	928	30.1	Not in COGs

The total is based on the total number of protein coding genes in the annotated genome.

Comparison with the genomes from other *Cellulomonas* species

Here, we compared the genome sequence of *C. massiliensis* strain JC225^T with those of *C. flavigena* strain 134^T [48] and *C. fimi* strain ATCC 484^T (EMBL accession number CP002666). The draft genome sequence of *C. massiliensis* has a smaller size than those of *C. flavigena* and *C. fimi* (3.40 vs 4.12 and 4.26 Mb, respectively), a lower G+C content (71.22 vs 74.3 and 74.7, respectively), and a smaller number of predicted genes (3,131 vs 3,788 and 3,863, respectively). In addition, *C. massiliensis* shared a mean 88.75% (range 70.01-100%) and 89.61% (range 70.07-100%) sequence similarity with *C. flavigena* and *C. fimi*, respectively, at the genome level.

Conclusion

On the basis of phenotypic, phylogenetic and genomic analyses, we formally propose the creation of *Cellulomonas massiliensis* sp. nov. that contains the strain JC225^T. This bacterium has been found in Senegal.

Description of *Cellulomonas massiliensis* sp. nov.

Cellulomonas massiliensis (ma.si.li.e'n.sis. L. gen. masc. n. *massiliensis*, of Massilia, the Latin name of Marseille where was isolated *C. massiliensis*).

Colonies are transparent and smooth with a diameter of 1 mm on blood-enriched Columbia agar and

Brain Heart Infusion (BHI) agar. Cells are rod-shaped with a diameter and length ranging from 0.37 to 0.60 µm (mean of 0.48 µm), and from 0.55 to 1.4 µm (mean of 0.95 µm), respectively. Optimal growth is achieved aerobically. Weak growth is observed with 5% CO₂ and under microaerophilic conditions. No growth is observed under anaerobic conditions. Growth occurs between 30-37°C, with optimal growth at 37°C. Cells stain Gram-positive, are non-endospore forming, and are motile. Catalase, oxidase, aesculin hydrolysis and β-galactosidase activities are present. Indole production, nitrate reduction, glucose fermentation, arginine dihydrolase, urease, gelatin hydrolysis, and glucose, arabinose, mannose, mannitol N-acetylglucosamine, maltose, gluconate, caprate, adipate, malate, citrate, and phenyl-acetate assimilation activities are absent. Cells are susceptible to amoxicillin, imipenem, ciprofloxacin and gentamicin, but resistant to trimethoprim/sulfamethoxazole and metronidazole. The 16S rRNA and genome sequences are deposited in Genbank and EMBL under accession numbers JN657218 and CAHD000000000, respectively. The G+C content of the genome is 71.22%. The type strain JC225^T (= CSUR P160 = DSM 25695) was isolated from the fecal flora of a healthy patient in Senegal.

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References

1. Lagier JC, Armougom F, Million M, Hugon P, Pagnier I, Robert C, Bittar F, Fournous G, Gimenez G, Maraninchi M, et al. Microbial culturomics: paradigm shift in the human gut microbiome study. *Clin Microbiol Infect* 2012; **18**:1185-1193. [PubMed](#)
2. Tindall BJ, Rossello-Mora R, Busse HJ, Ludwig W, Kämpfer P. Notes on the characterization of prokaryote strains for taxonomic purposes. *Int J Syst Evol Microbiol* 2010; **60**:249-266. [PubMed](#) <http://dx.doi.org/10.1099/ijs.0.016949-0>
3. Genome Online Database. <http://www.genomesonline.org/cgi-bin/GOLD/index.cgi>
4. Welker M, Moore ER. Applications of whole-cell matrix-assisted laser-desorption/ionization time-of-flight mass spectrometry in systematic microbiology. *Syst Appl Microbiol* 2011; **34**:2-11. [PubMed](#) <http://dx.doi.org/10.1016/j.syapm.2010.11.013>
5. Lagier JC, El Karkouri K, Nguyen TT, Armougom F, Raoult D, Fournier PE. Non-contiguous finished genome sequence and description of *Anaerococcus senegalensis* sp. nov. *Stand Genomic Sci* 2012; **6**:116-125. [PubMed](#) <http://dx.doi.org/10.4056/sigs.2415480>
6. Kokcha S, Mishra AK, Lagier JC, Million M, Leroy Q, Raoult D, Fournier PE. Non contiguous-finished genome sequence and description of *Bacillus timonensis* sp. nov. *Stand Genomic Sci* 2012; **6**:346-355. <http://dx.doi.org/10.4056/sigs.2776064>

7. Mishra AK, Gimenez G, Lagier JC, Robert C, Raoult D, Fournier PE. Non-contiguous finished genome sequence and description of *Alistipes senegalensis* sp. nov. *Stand Genomic Sci* 2012; **6**:304-314. <http://dx.doi.org/10.4056/sigs.2625821>
8. Lagier JC, Armougom F, Mishra AK, Ngyuen TT, Raoult D, Fournier PE. Non-contiguous finished genome sequence and description of *Alistipes timonensis* sp. nov. *Stand Genomic Sci* 2012; **6**:315-324. <http://dx.doi.org/10.4056/sigs.2685917>
9. Mishra AK, Lagier JC, Robert C, Raoult D, Fournier PE. Non-contiguous finished genome sequence and description of *Clostridium senegalense* sp. nov. *Stand Genomic Sci* 2012; **6**:386-395.
10. Mishra AK, Lagier JC, Robert C, Raoult D, Fournier PE. Non-contiguous finished genome sequence and description of *Peptoniphilus timonensis* sp. nov. *Stand Genomic Sci* 2012; **7**:1-11. <http://dx.doi.org/10.4056/sigs.2956294>
11. Mishra AK, Lagier JC, Rivet R, Raoult D, Fournier PE. Non-contiguous finished genome sequence and description of *Paenibacillus senegalensis* sp. nov. *Stand Genomic Sci* 2012; **7**:70-81. <http://dx.doi.org/10.4056/sigs.3054650>
12. Bergey DH, Harrison FC, Breed RS, Hammer BW, Huntton FM. In: Bergey DH, Harrison FC, Breed RS, Hammer BW, Huntton FM (eds), Bergey's Manual of Determinative Bacteriology, First Edition, The Williams and Wilkins Co., Baltimore, 1923, p. 1-442.
13. Lee CM, Weon HY, Hong SB, Jeon YA, Schumann P, Kroppenstedt RM, Kwon SW, Stackebrandt E. *Cellulomonas aerilata* sp. nov., isolated from an air sample. *Int J Syst Evol Microbiol* 2008; **58**:2925-2929. <http://dx.doi.org/10.1099/ijs.0.2008/002253-0>
14. Jones BE, Grant WD, Duckworth AW, Schumann P, Weiss N, Stackebrandt E. *Cellulomonas bogoriensis* sp. nov., an alkaliphilic cellulomonad. *Int J Syst Evol Microbiol* 2005; **55**:1711-1714. <http://dx.doi.org/10.1099/ijs.0.63646-0>
15. Shi Z, Luo G, Wang G. *Cellulomonas carbonis* sp. nov., isolated from coal mine soil. *Int J Syst Evol Microbiol* 2012; **62**:2004-2010. <http://dx.doi.org/10.1099/ijs.0.034934-0>
16. Yoon MH, Ten LN, Im WT, Lee ST. *Cellulomonas chitinilytica* sp. nov., a chitinolytic bacterium isolated from cattle-farm compost. *Int J Syst Evol Microbiol* 2008; **58**:1878-1884. [PubMed](http://dx.doi.org/10.1099/ijs.0.64768-0) <http://dx.doi.org/10.1099/ijs.0.64768-0>
17. Kang MS, Im WT, Jung HM, Kim MK, Goodfellow M, Kim KK, Yang HC, An DS, Lee ST. *Cellulomonas composti* sp. nov., a cellulolytic bacterium isolated from cattle farm compost. *Int J Syst Evol Microbiol* 2007; **57**:1256-1260. [PubMed](http://dx.doi.org/10.1099/ijs.0.63974-0) <http://dx.doi.org/10.1099/ijs.0.63974-0>
18. Brown JM, Frazier RP, Morey RE, Steigerwalt AG, Pellegrini GJ, Daneshvar MI, Hollis DG, McNeil MM. Phenotypic and genetic characterization of clinical isolates of CDC coryneform group A-3: proposal of a new species of *Cellulomonas*, *Cellulomonas denverensis* sp. nov. *J Clin Microbiol* 2005; **43**:1732-1737. [PubMed](http://dx.doi.org/10.1128/JCM.43.4.1732-1737.2005) <http://dx.doi.org/10.1128/JCM.43.4.1732-1737.2005>
19. Funke G, Ramos CP, Collins MD. Identification of some clinical strains of CDC coryneform group A-3 and A-4 bacteria as *Cellulomonas* species and proposal of *Cellulomonas hominis* sp. nov. for some group A-3 strains. *J Clin Microbiol* 1995; **33**:2091-2097. [PubMed](http://dx.doi.org/10.1099/ijs.0.2008/002253-0)
20. Collins MD, Pascual C. Reclassification of *Actinomyces humiferus* (Gledhill and Casida) as *Cellulomonas humilata* nom. corrig., comb. nov. *Int J Syst Evol Microbiol* 2000; **50**:661-663. [PubMed](http://dx.doi.org/10.1099/00207713-50-2-661) <http://dx.doi.org/10.1099/00207713-50-2-661>
21. Elberson MA, Malekzadeh F, Yazdi MT, Kameranpour N, Noori-Dalooi MR, Matte MH, Shahamat M, Colwell RR, Sowers KR. *Cellulomonas persica* sp. nov. and *Cellulomonas iranensis* sp. nov., mesophilic cellulose-degrading bacteria isolated from forest soils. *Int J Syst Evol Microbiol* 2000; **50**:993-996. [PubMed](http://dx.doi.org/10.1099/00207713-50-3-993) <http://dx.doi.org/10.1099/00207713-50-3-993>
22. Rusznyák A, Toth EM, Schumann P, Sproer C, Makk J, Szabo G, Vladar P, Marialigeti K, Borsodi AK. *Cellulomonas phragmiteti* sp. nov., a cellulolytic bacterium isolated from reed (*Phragmites australis*) periphyton in a shallow soda pond. *Int J Syst Evol Microbiol* 2011; **61**:1662-1666. [PubMed](http://dx.doi.org/10.1099/ijs.0.022608-0) <http://dx.doi.org/10.1099/ijs.0.022608-0>
23. An DS, Im WT, Yang HC, Kang MS, Kim KK, Jin L, Kim MK, Lee ST. *Cellulomonas terrae* sp. nov., a cellulolytic and xylanolytic bacterium isolated from soil. *Int J Syst Evol Microbiol* 2005; **55**:1705-1709. [PubMed](http://dx.doi.org/10.1099/ijs.0.63696-0) <http://dx.doi.org/10.1099/ijs.0.63696-0>
24. Rivas R, Trujillo ME, Mateos PF, Martinez-Molina E, Velazquez E. *Cellulomonas xylanilytica* sp.

- nov., a cellulolytic and xylanolytic bacterium isolated from a decayed elm tree. *Int J Syst Evol Microbiol* 2004; **54**:533-536. [PubMed](#) <http://dx.doi.org/10.1099/ijs.0.02866-0>
25. Lai PC, Chen YS, Lee SS. Infective endocarditis and osteomyelitis caused by *Cellulomonas*: a case report and review of the literature. *Diagn Microbiol Infect Dis* 2009; **65**:184-187. [PubMed](#) <http://dx.doi.org/10.1016/j.diagmicrobio.2009.06.001>
 26. Sharma S, Saffra NA, Chinyadza T, Ghitan M, Chapnick EK. Endocapsular cellulomonas as a cause of persistent postoperative endophthalmitis. *Ophthalmic Surg Lasers Imaging* 2008; **39**:328-330. [PubMed](#) <http://dx.doi.org/10.3928/15428877-20080701-16>
 27. Ohtaki H, Ohkusu K, Sawamura H, Ohta H, Inoue R, Iwasa J, Ito H, Murakami N, Ezaki T, Moriwaki H, Seishima M. First report of acute cholecystitis with sepsis caused by *Cellulomonas denverensis*. *J Clin Microbiol* 2009; **47**:3391-3393. [PubMed](#) <http://dx.doi.org/10.1128/JCM.00678-09>
 28. Trape JF, Tall A, Diagne N, Ndiath O, Ly AB, Faye J, Dieye-Ba F, Roucher C, Bouganali C, Badiane A, et al. Malaria morbidity and pyrethroid resistance after the introduction of insecticide-treated bednets and artemisinin-based combination therapies: a longitudinal study. *Lancet Infect Dis* 2011; **11**:925-932. [PubMed](#) [http://dx.doi.org/10.1016/S1473-3099\(11\)70194-3](http://dx.doi.org/10.1016/S1473-3099(11)70194-3)
 29. Woese CR, Kandler O, Wheelis ML. Towards a natural system of organisms: proposal for the domains *Archaea*, *Bacteria*, and *Eucarya*. *Proc Natl Acad Sci USA* 1990; **87**:4576-4579. [PubMed](#) <http://dx.doi.org/10.1073/pnas.87.12.4576>
 30. Garrity GM, Holt JG. The Road Map to the Manual. In: Garrity GM, Boone DR, Castenholz RW (eds), *Bergey's Manual of Systematic Bacteriology*, Second Edition, Volume 1, Springer, New York, 2001, p. 119-169.
 31. Stackebrandt E, Ebers J, Rainey F, Ward-Rainey N. Proposal for a New Hierarchic Classification System, *Actinobacteria* classis nov. *Int J Syst Bacteriol* 1997; **47**:479-491. [PubMed](#) <http://dx.doi.org/10.1099/00207713-47-2-479>
 32. Skerman V, McGowan V, Sneath P. Approved Lists of Bacterial Names. *Int J Syst Bacteriol* 1980; **30**:225-420. [PubMed](#) <http://dx.doi.org/10.1099/00207713-30-1-225>
 33. Buchanan RE. Studies in the nomenclature and classification of bacteria. II. The primary subdivisions of the *Schizomycetes*. *J Bacteriol* 1917; **2**:155-164. [PubMed](#)
 34. Zhi XY, Li WJ, Stackebrandt E. An update of the structure and 16S rRNA gene sequence-based definition of higher ranks of the class *Actinobacteria*, with the proposal of two new suborders and four new families and emended descriptions of the existing higher taxa. *Int J Syst Evol Microbiol* 2009; **59**:589-608. [PubMed](#) <http://dx.doi.org/10.1099/ijs.0.65780-0>
 35. Stackebrandt E, Prauser H. Assignment of the genera *Cellulomonas*, *Oerskovia*, *Promicromonospora*, and *Jonesia* to *Cellulomonadaceae* fam. nov. *Syst Appl Microbiol* 1991; **14**:261-265. [PubMed](#) [http://dx.doi.org/10.1016/S0723-2020\(11\)80378-4](http://dx.doi.org/10.1016/S0723-2020(11)80378-4)
 36. List Editor. Validation of the publication of new names and new combinations previously effectively published outside the IJSB. List no. 39. *Int J Syst Bacteriol* 1991; **41**:580-581. [PubMed](#) <http://dx.doi.org/10.1099/00207713-41-4-580>
 37. Stackebrandt E, Schumann P. Description of *Bogoriellaceae* fam. nov., *Dermaococcaceae* fam. nov., *Rarobacteraceae* fam. nov. and *Sanguibacteraceae* fam. nov. and emendation of some families of the suborder *Micrococcineae*. *Int J Syst Evol Microbiol* 2000; **50**:1279-1285. [PubMed](#) <http://dx.doi.org/10.1099/00207713-50-3-1279>
 38. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet* 2000; **25**:25-29. [PubMed](#) <http://dx.doi.org/10.1038/75556>
 39. Stackebrandt E, Ebers J. Taxonomic parameters revisited: tarnished gold standards. *Microbiol Today* 2006; **33**:152-155.
 40. GenBank database. <http://www.ncbi.nlm.nih.gov/genbank>
 41. Seng P, Drancourt M, Gouriet F, La Scola B, Fournier PE, Rolain JM, Raoult D. Ongoing revolution in bacteriology: routine identification of bacteria by matrix-assisted laser desorption ionization time-of-flight mass spectrometry. *Clin Infect Dis* 2009; **49**:543-551. [PubMed](#) <http://dx.doi.org/10.1086/600885>
 42. Field D, Garrity G, Gray T, Morrison N, Selengut J, Sterk P, Tatusova T, Thomson N, Allen MJ,

- Angiuoli SV, et al. The minimum information about a genome sequence (MIGS) specification. *Nat Biotechnol* 2008; **26**:541-547. [PubMed](#) <http://dx.doi.org/10.1038/nbt1360>
43. Prodigal. <http://prodigal.ornl.gov>
44. Lowe TM, Eddy SR. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 1997; **25**:955-964. [PubMed](#)
45. Lagesen K, Hallin P, Rodland EA, Staerfeldt HH, Rognes T, Ussery DW. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 2007; **35**:3100-3108. [PubMed](#) <http://dx.doi.org/10.1093/nar/gkm160>
46. Krogh A, Larsson B, von HG, Sonnhammer EL. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 2001; **305**:567-580. [PubMed](#) <http://dx.doi.org/10.1006/jmbi.2000.4315>
47. Bendtsen JD, Nielsen H, von HG, Brunak S. Improved prediction of signal peptides: SignalP 3.0. *J Mol Biol* 2004; **340**:783-795. [PubMed](#) <http://dx.doi.org/10.1016/j.jmb.2004.05.028>
48. Abt B, Foster B, Lapidus A, Clum A, Sun H, Pukall R, Lucas S, Glavina Del Rio T, Nolan M, Tice H, et al. Complete genome sequence of *Cellulomonas flavigena* type strain (134^T). *Stand Genomic Sci* 2010; **3**:15-25. [PubMed](#) <http://dx.doi.org/10.4056/sigs.1012662>
49. Cavalier-Smith T. The neomuran origin of archaeobacteria, the negibacterial root of the universal tree and bacterial megaclassification. *Int J Syst Evol Microbiol* 2002; **52**:7-76. [PubMed](#)